

ITS-Animal-Free[™] Supplementation for iPSC Expansion and Pluripotency: A Platform Approach for Serum-Free Cell Expansion

Mark Stathos, PhD, and Andrew Hamann, PhD InVitria, Inc., USA

APPLICATION BENEFITS

- ITS Animal-Free[™] is an essential serum replacement – Contains Insulin, Transferrin, and Selenium, core components of E8 media critical for iPSC culture
- Enhances iPSC proliferation Supports rapid expansion in serum-free and chemically defined media
- Maintains pluripotency Sustains high Oct4, Sox2, and Nanog expression during expansion
- Ready-to-use, animal-origin-free

 Pre-optimized 100x liquid
 format simplifies custom media
 formulation and supports
 regulatory compliance

INVITRIA SOLUTIONS

<u>ITS Animal-Free</u> - Blood-free cell culture media supplement

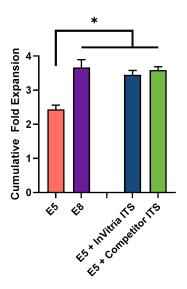


EXECUTIVE SUMMARY

ITS Animal-Free (ITS AF), InVitria's 100x cocktail of Insulin, Transferrin, and Selenium that is completely animal-origin-free, is essential for culture of all mammalian cell types including iPSCs. ITS AF can be conveniently supplemented into custom media formulations to replace the core components of serum. This reduces variability, eliminates safety and regulatory hurdles, and allows users to focus on the finer points when optimizing custom media formulations. Here we show that InVitria[®] ITS AF enhances iPSC proliferation and maintains pluripotency comparably to the leading competitors.

Key Highlights

- · Enables robust iPSC proliferation in serum-free systems
- Maintains ≥85% Oct4+/Sox2+/Nanog+ pluripotent population
- · Performs equivalently to leading ITS products without optimization
- Animal-origin-free and blood-free, supporting GMP-ready workflows



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Figure 1. ITS AF significantly increases proliferation of iPSCs and performs comparably to the leading competitor at a range of concentrations.

Figure 2. ITS AF significantly enhances pluripotency of human iPSC cultures comparably to or better than a leading competitor product.

INTRODUCTION

Since the discovery of induced pluripotent stem cells (iPSCs) (Takahashi and Yamanaka, 2006), scientists have sought to better define culture conditions for this highly sensitive cell type to enable facile culture, maintenance of pluripotency, and controlled reproducible differentiation into many target cell types (Villa-Diaz et al., 2013). The earliest stem cell protocols required feeder cells and serum (Thomson et al., 1998). The feeder cells initially employed introduced significant variability and required complex workflows (Li et al. 2017) and serum inclusion was also untenable due to the risk of xenogeneic contamination, regulatory concerns, and variability due to undefined components such as growth factors that induced unwanted differentiation (Lyra-Leite et al., 2022).

To address this, members of James Thomson's lab developed the fully defined minimal essential iPSC media, Essential 8[™] (E8) (Chen et al. 2011). This formulation consists of just eight ingredients including a base media (DMEM/F12), a buffering agent (sodium bicarbonate), an antioxidant (vitamin C), two growth factors (TGFβ and FGF2) as well as Insulin, Transferrin, and Selenium (ITS). It is no coincidence that ITS was included in E8 media as these components have all long been known to play indispensable roles in cell culture and to act in concert as a serum replacement (Guilbert and Iscove, 1976).

Insulin has been demonstrated to be a universal growth factor in human cell culture (Barnes and Sato, 1980). It activates cell signaling pathways that drive cell cycle progression even in cells that are arrested in G0/G1 due to nutrient limitation and plays important roles in cell metabolism (Straus, 1981).

Transferrin is the principle physiological delivery vehicle of iron to cells. Iron is essential for cellular respiration (Luck and Mason, 2015) and is a crucial co-factor for enzymes involved in DNA synthesis (Laskey, 1988). Furthermore, transferrin delivers iron through receptor mediated endocytosis, meaning cells can regulate its intake to minimize iron-induced oxidative stress.

Selenium is a trace element that is incorporated into antioxidant enzymes like glutathione peroxidase and thioredoxin reductase in the form of the noncanonical amino acid selenocysteine. It is essential for moderating oxidative stress and plays a critical role in cell cycle progression (Zeng, 2009).

InVitria's ITS-AF is a pre-formulated, optimized, animal-origin-free serum replacement cocktail conveniently available as a 100x liquid solution. This format not only enables serum replacement for all cell types but also facilitates custom media formulation which is ubiquitous among clinical iPSC research groups.

Here we showcase directly the critical role of ITS in proliferation and maintenance of pluripotency in iPSCs. Due to the essential nature of ITS components, these results are applicable to any human cell type.

RESULTS AND DISCUSSION

ITS Enhances iPSC Proliferation

To assess how ITS supplementation affects iPSC proliferation, researchers first created an in-house "E5" medium—identical to commercial E8 but lacking ITS. InVitria ITS-AF or a leading competitor's ITS was then added back into E5 at final concentrations ranging from 1x to 5x. (For reference, standard E8 media contains 2x ITS.)

Human iPSCs previously maintained in E8 were plated in 96-well plates with 10 μ M Y-27632 ROCK inhibitor. After 24 hours, cells were switched to one of the following conditions: E5 alone, E5 with InVitria ITS-AF, E5 with competitor ITS, or complete E8. Cell proliferation was monitored every 2 hours over the next 48 hours using high-content imaging.

As shown in **Figure 1**, both InVitria ITS AF and competitor ITS supported robust expansion at 2xconcentrations, equivalent to E8. Results at other concentrations (1x-5x) showed similar trends and are omitted for clarity. The consistency across doses indicates that InVitria ITS AF does not require titration or optimization during media formulation.

All media conditions containing ITS supported significantly greater iPSC proliferation compared to the E5 control lacking ITS. This difference is likely due to the absence of insulin (which supports metabolic activity), transferrin (which delivers iron for respiration and DNA synthesis), and selenium (which protects against oxidative stress).

APPLICATION NOTE

No significant differences were observed between the InVitria ITS AF and the competitor formulations at any tested concentration, indicating comparable performance. These findings suggest that ITS-AF does not require further optimization during media development. Given the essential roles of ITS components, it is expected that these benefits would become even more pronounced over extended culture periods.

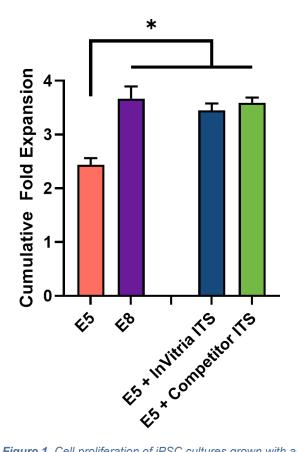


Figure 1. Cell proliferation of iPSC cultures grown with and without InVitria ITS AF or competitor ITS. Human iPSCs were maintained for 48 hours in E5 media without ITS, off the shelf E8 media, or E5 media with 2x ITS from InVitria or a competitor (equivalent to E8 ITS levels). Cumulative fold expansion was measured 48 hours after plating. All formulations containing ITS yielded cell populations that were significantly more pluripotent than the E5 negative control (One Way ANOVA with Tukey post hoc test, p < 0.05).

ITS Maintains Pluripotency in iPSCs

To further characterize the effect of ITS on iPSC culture, the cells that had been expanded in different amounts of ITS were then analyzed by flow cytometry for the core pluripotency markers Oct4, Sox2, and Nanog which are essential for maintenance of iPSCs (Rodda, 2005) (**Figure 2**).

All conditions supplemented with ITS maintained at least 85% triple-positive cells, indicating high pluripotency. In contrast, the ITS-free control group showed only 55% pluripotency. The larger variability in this negative control is likely due to the limited number of viable cells available for analysis. The observed loss of pluripotency may be linked to disrupted iron homeostasis in the absence of transferrin (Han et al., 2018).

Taken together with the expansion data, this phenotyping result suggests that ITS is critical for maintaining proliferating cells as well as preventing iPSCs from spontaneously differentiating. Once again there were no significant differences between conditions that contained ITS suggesting that InVitria's ITS-AF is comparable to the leading competitor at every dose evaluated. Furthermore, the lack of a dose response suggests that ITS does not need to be titrated which simplifies custom media formulation.

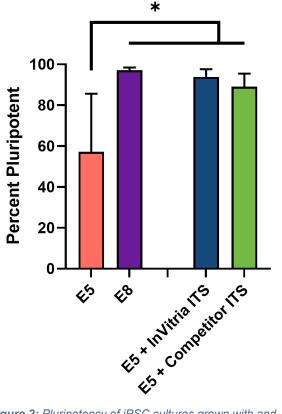


Figure 2: Pluripotency of iPSC cultures grown with and without InVitria ITS AF or competitor ITS. Human iPSCs were maintained for 48 hours in E5 media, off the shelf E8 or E5 plus 2x, ITS from InVitria or a competitor (equivalent ITS levels to E8). Pluripotency was defined by Oct4, Sox2, and Nanog triple positive staining measured by intracellular flow cytometry. All formulations containing ITS yielded cell populations that were significantly more pluripotent than the E5 negative control (One Way ANOVA with Tukey post hoc test, p < 0.05).

MATERIALS & METHODS

Human iPSCs maintained in Essential 8 media (ThermoFisher Scientific) were dissociated using Accutase[®] (ThermoFisher Scientific) and seeded into 96-well plates at 10,000 cells per well. Each well was coated with 0.5 µg/cm² N-terminal truncated Vitronectin[™] (ThermoFisher Scientific) and supplemented with 10 µM Y-27632 ROCK inhibitor. Cells were cultured in one of the following conditions: E5 media (homemade Essential 8 without ITS), E5 supplemented with 1–5x InVitria ITS AF or competitor ITS, or complete commercial E8. Plates were incubated on an Incucyte SX5 system, and proliferation was monitored via confluence measurements taken every 2 hours. After 24 hours, the media was changed to remove ROCKi, and monitoring continued for an additional 24 hours.

At 48 hours, cells were harvested using Accutase and stained for flow cytometry. Viability was assessed using <u>Zombie Aqua</u>[™] fixable dye (<u>Biolegend</u>). Cells were fixed and permeabilized with the <u>FOXP3 eBioscience</u>[™] buffer set (ThermoFisher Scientific), then stained with pluripotency markers: Oct4 <u>Alexa Fluor® 647</u>, <u>Sox2 Alexa Fluor</u> <u>488</u>, and <u>Nanog Alexa Fluor 594</u> (Biolegend). Samples were analyzed using a <u>CytoFLEX LX</u>[™] flow cytometer (<u>Beckman Coulter</u>). Pluripotency was defined as the percentage of live cells triple-positive for Oct4, Sox2, and Nanog.

CONCLUSION

This dataset demonstrates that ITS is essential for maintaining both expansion and pluripotency of iPSC cultures. Without these critical components, cultures fail to propagate efficiently and begin to spontaneously differentiate. Additionally, InVitria ITS-AF was shown perform as well as a leading competitor ITS product while also being completely defined and animal free. This makes InVitria ITS-AF a strong foundation on which to build custom media formulations that are high performance, chemically defined, and animal free.

REFERENCES

- Takahashi, K., & Yamanaka, S. (2006). Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. Cell, 126(4), 663–676. <u>https://doi.org/10.1016/j.cell.2006.07.024</u>
- Villa-Diaz, L. G., Ross, A. M., Lahann, J., & Krebsbach, P. H. (2013). Concise review: The evolution of human pluripotent stem cell culture: From feeder cells to synthetic coatings. Stem Cells, 31(1), 1–7. <u>https://doi.org/10.1002/stem.1190</u>
- Thomson, J. A., Itskovitz-Eldor, J., Shapiro, S. S., Waknitz, M. A., Swiergiel, J. J., Marshall, V. S., & Jones, J. M. (1998). Embryonic stem cell lines derived from human blastocysts. Science, 282(5391), 1145–1147. <u>https://doi.org/10.1126/science.282.5391.1145</u>
- Li, P., Wang, S., Zhan, L., He, X., Chi, G., Lv, S., Xu, Z., Xia, Y., Teng, S., Li, L., & Li, Y. (2017). Efficient feeder cells preparation system for large-scale preparation and application of induced pluripotent stem cells. Scientific Reports, 7, 12266. https://doi.org/10.1038/s41598-017-10428-5
- Lyra-Leite, D. M., Gutiérrez-Gutiérrez, Ó., Wang, M., Zhou, Y., Cyganek, L., & Burridge, P. W. (2022). A review of protocols for human iPSC culture, cardiac differentiation, subtype-specification, maturation, and direct reprogramming. STAR Protocols, 3(3), 101560. <u>https://doi.org/10.1016/j.xpro.2022.101560</u>
- Chen, G., Gulbranson, D. R., Hou, Z., Bolin, J. M., Ruotti, V., Probasco, M. D., Smuga-Otto, K., Howden, S. E., Diol, N. R., Propson, N. E., & Thomson, J. A. (2011). Chemically defined conditions for human iPSC derivation and culture. Nature Methods, 8(5), 424–429. <u>https://doi.org/10.1038/nmeth.1593</u>
- Guilbert, L. J., & Iscove, N. N. (1976). Partial replacement of serum by transferrin, albumin and lecithin in haemopoietic cell cultures. Nature, 263(5574), 594–595. <u>https://doi.org/10.1038/263594a0</u>
- Barnes, D., & Sato, G. (1980). Serum-free cell culture: A unifying approach. Cell, 22(3), 649–655. https://doi.org/10.1016/0092-8674(80)90540-1
- Straus, D. S. (1981). Effects of insulin on cellular growth and proliferation. Life Sciences, 29(21), 2131–2139. https://doi.org/10.1016/0024-3205(81)90482-3
- Luck, A. N., & Mason, A. B. (2012). Transferrin-mediated cellular iron delivery. Current Topics in Membranes, 69, 3–35. https://doi.org/10.1016/B978-0-12-394390-3.00001-X

APPLICATION NOTE

- Laskey, J., Webb, I., Schulman, H. M., & Ponka, P. (1988). Evidence that transferrin supports cell proliferation by supplying iron for DNA synthesis. Experimental Cell Research, 176(1), 87–95. <u>https://doi.org/10.1016/0014-4827(88)90123-1</u>
- Zeng, H. (2009). Selenium as an essential micronutrient: Roles in cell cycle and apoptosis. Molecules, 14(3), 1263–1278. https://doi.org/10.3390/molecules14031263
- Rodda, D. J., Chew, J. L., Lim, L. H., Loh, Y. H., Wang, B., Ng, H. H., & Robson, P. (2005). Transcriptional regulation of Nanog by OCT4 and SOX2. Journal of Biological Chemistry, 280(26), 24731–24737. <u>https://doi.org/10.1074/jbc.M502573200</u>
- Han, Z., Yu, Y., Xu, J., Bao, Z., Xu, Z., Hu, J., Yu, M., Bamba, D., Ma, W., Ding, F., Zhang, L., Jin, M., Yan, G., Huang, Q.,
 Wang, X., Hua, B., Yang, F., Li, Y., Lei, L., ... Cai, B. (2019). Iron homeostasis determines fate of human pluripotent stem cells via glycerophospholipids-epigenetic circuit. Stem Cells, 37(4), 489–503. <u>https://doi.org/10.1002/stem.2967</u>

ORDERING INFORMATION

ITS Animal-Free - Blood-free cell culture media supplement	<u>777ITS091</u>
ITS Animal-Free- 10 mL Bottle	777ITS091-10mL
ITS Animal-Free- 100 mL Bottle	777ITS091-100mL
ITS Animal-Free- 10 x 10 mL Bottles	777ITS091-10mLx10

ABOUT INVITRIA

InVitria is a global leader in developing and manufacturing high-performance, animal-origin-free cell culture supplements and recombinant proteins. Our products eliminate human and animal serum-derived components to improve consistency, reduce regulatory burden, and accelerate the approval of life-changing therapies. Trusted in clinical and commercial production of cell and gene therapies, vaccines, regenerative medicine, and medical devices, InVitria supports cGMP and ISO-compliant manufacturing aligned with FDA and EMA expectations. Scalable to meet growing global demand, our chemically defined solutions enable safer, more efficient biomanufacturing.



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InVitria, Inc.

2718 Industrial Drive Junction City, KS 66441 USA Phone: 800-916-8311 Fax: 785-238-5688 Email: info@invitria.com www.invitria.com